

## BBA Report

---

BBA 71482

### VANADATE-INDUCED INHIBITION OF SODIUM TRANSPORT AND OF SODIUM-INDEPENDENT ANION TRANSPORT IN TURTLE BLADDER

GERHARD EHRENSPECK

*Department of Zoology and Microbiology and College of Osteopathic Medicine, Ohio University, Athens, OH 45701 (U.S.A.)*

(Received March 28th, 1980)

*Key words: Vanadate inhibition; Na<sup>+</sup> transport; Anion transport; Cl<sup>-</sup> reabsorption; Electrical phenomenon; (Turtle bladder)*

#### Summary

Vanadate in the serosal bathing fluid of turtle bladders inhibits the Na<sup>+</sup> moiety of the short-circuiting current ( $I_{sc}$ ), the anion ( $Cl^-$ ,  $HCO_3^-$ ) moiety of  $I_{sc}$ , and net  $Cl^-$  flux. Since the anion transport is Na<sup>+</sup>-independent and ouabain-insensitive, its inhibition by vanadate is uniquely different from the well known vanadate-induced inhibition of (Na<sup>+</sup>+K<sup>+</sup>)-ATPase and Na<sup>+</sup> transport-dependent anion movement of some other epithelia. Vanadate also generates damped oscillations in the bladders' electrical parameters, an unusual effect by an ion in epithelial systems.

---

Vanadate (V in the +5 oxidation state) possesses a striking natriuretic and diuretic action [1], presumably related to its inhibition of both (Na<sup>+</sup>+K<sup>+</sup>)-ATPase [2, 3] and vasopressin-sensitive, cyclic AMP-dependent Na<sup>+</sup> and water transport [4]. The recent report by Cantley et al. [5] that vanadate enters the red cell via a disulfonic stilbene-sensitive pathway suggested to me that this oxyanion might interfere with anion transport in the turtle bladder, a well known model epithelium that possesses disulfonic stilbene-inhibitable anion-selective pathways [6–8]. The present report describes some properties of the vanadate-induced inhibition of anion and Na<sup>+</sup> transport across bladders of *Pseudemys scripta* turtles.

Methods for evaluating transepithelial potential ( $PD$ ), short-circuiting current ( $I_{sc}$ ), d.c. resistance ( $R$ ), and  $^{36}Cl^-$  fluxes have been described [7, 9]. For studies of Na<sup>+</sup> transport, paired hemi-bladders were bathed on both surfaces by identical Na<sup>+</sup> Ringer solutions; for studies on anion transport, 0.2 mM ouabain was present in the serosal fluid. Sodium metavanadate (Fisher-

Scientific) was used at concentrations of 0.1 and 1 mM. Under the experimental conditions used, the predominant anionic species of vanadate presumably was  $\text{H}_2\text{VO}_4^-$  [10].

Table I shows the effect of vanadate on 13 bladders bathed by  $\text{Na}^+$  media without exogenous  $\text{HCO}_3^-$ .  $I_{\text{sc}}$ , which under these bathing conditions approximates net reabsorption of  $\text{Na}^+$  [11–13], was decreased by 65.8% in 2 h. Not shown is that the onset of inhibition consistently occurred within 2–5 min after vanadate addition, but that the rate of decline in the measured parameters varied widely (half-time of  $84 \pm 13$  min). No inhibition of  $I_{\text{sc}}$  was observed at 0.1 mM vanadate. In five control bladders the time-dependent decay in  $I_{\text{sc}}$  was  $4.6 \pm 1.4\%$  per h.

TABLE I

EFFECT OF VANADATE ON  $PD$ ,  $I_{\text{sc}}$  AND  $R$  OF BLADDERS IN  $\text{HCO}_3^-$ -FREE  $\text{Na}^+$  BATHING MEDIUM

Mean values  $\pm$  S.E. ( $n = 13$ ) of  $PD$ ,  $I_{\text{sc}}$ , and  $R$  and of the percentage changes in these parameters 2 h after serosal addition of 1 mM vanadate. Positive values of  $PD$  and  $I_{\text{sc}}$  indicate that serosa is electro-positive to mucosa. Area of exposed tissue,  $1.5 \text{ cm}^2$ . Composition of bathing fluid (mM):  $\text{NaCl}$ , 83.5;  $\text{Na}_2\text{SO}_4$ , 10.0;  $\text{KCl}$ , 4;  $\text{MgSO}_4$ , 0.8;  $\text{CaSO}_4$ , 2.0;  $\text{K}_2\text{HPO}_4$ , 0.65;  $\text{KH}_2\text{PO}_4$ , 0.10; glucose, 11; osmolality was adjusted to 220 mosM/kg with sucrose; final pH,  $7.6 \pm 0.1$ ; aspirated with  $\text{H}_2\text{O}$ -saturated 100%  $\text{O}_2$ .

Condition	$PD$ (mV)	$I_{\text{sc}}$ ( $\mu\text{A}$ )	$R$ ( $\text{k}\Omega$ )
Before vanadate	$67.5 \pm 4.1$	$108.0 \pm 8.1$	$0.56 \pm 0.04$
After vanadate	$21.6 \pm 3.3$	$40.2 \pm 6.2$	$0.58 \pm 0.06$
$\Delta(\%)^*$	$-69.6 \pm 4.3$	$-65.8 \pm 4.5$	$-1.4 \pm 9.1$
$P(\Delta=0)$	$P < 0.001$	$P < 0.001$	$P > 0.8$

\*Note that values of  $\Delta(\%)$  are means  $\pm$  S.E. of  $n$  individual percentage changes in the designated electrical parameters after addition of vanadate. Statistical significance of  $\Delta(\%)$  was calculated by the Student's  $t$ -test.

Table II shows the effect of vanadate on  $PD$ ,  $I_{\text{sc}}$ , and  $R$  of 14 bladders (seven pairs of mated hemi-bladders) bathed by  $\text{Na}^+(\text{Cl}^- + \text{HCO}_3^-)$  media plus serosal ouabain.  $I_{\text{sc}}$ , which under these bathing conditions is reversed in direction and approximates the algebraic sum of the conductive flows of  $\text{Cl}^-$  and  $\text{HCO}_3^-$  [12, 14, 15], was decreased by 72.2% in 2 h. Other results common to both  $\text{Na}^+$ -dependent (Table I) and ouabain-insensitive moieties (Table II) of  $PD$  and  $I_{\text{sc}}$  were the following: (i) washing of the serosal surface or exposure to 2 mM norepinephrine [2] failed to restore these parameters to control values; (ii) 1 mM vanadate in the mucosal fluid for at least 1 h failed to decrease  $PD$  and  $I_{\text{sc}}$ .

Because of the possibility of a non-conductive  $\text{Cl}^-$ - $\text{HCO}_3^-$  exchange component of  $\text{Cl}^-$  transport [16, 17] which would not contribute to  $I_{\text{sc}}$ ,  $^{36}\text{Cl}^-$  fluxes were measured in five of the mated pairs of hemi-bladders of Table II. The measured and calculated  $\text{Cl}^-$  flux parameters are shown in Table III. The major effect was a 53.7% decrease in the M-to-S flux (M, mucosa; S, serosa). Since the S-to-M fluxes were low relative to the M-to-S fluxes, the changes in the S-to-M fluxes contributed comparatively little to the changes in the net M-to-S  $\text{Cl}^-$  flux. The effects of time-dependent

TABLE II

EFFECT OF VANADATE ON  $PD$ ,  $I_{sc}$ , AND  $R$  OF OUABAIN-TREATED BLADDERS IN  $Na^+$  ( $Cl^- + HCO_3^-$ ) BATHING MEDIUM

Mean values  $\pm$  S.E. ( $n = 14$ , seven mated pairs of hemi-bladders) of  $PD$ ,  $I_{sc}$ , and  $R$  and of percentage changes in these parameters 2 h after serosal addition of 1 mM vanadate. Negative values of  $PD$  and  $I_{sc}$  indicate that serosa is electronegative to mucosa. Area of exposed tissue,  $1.5 \text{ cm}^2$ . Composition of bathing fluid (mM):  $NaCl$ , 21;  $NaHCO_3$ , 20;  $Na_2SO_4$ , 30;  $KCl$ , 4;  $MgSO_4$ , 0.8;  $CaSO_4$ , 2.0;  $K_2HPO_4$ , 0.65;  $KH_2PO_4$ , 0.1; glucose, 11; osmolality was adjusted to 220 mosM/kg with sucrose; final pH  $7.6 \pm 0.1$ ; aspirated with  $H_2O$ -saturated 98%  $O_2$ /2%  $CO_2$ ; 0.2 mM ouabain in serosal fluid. Statistical definitions given in Table I.

Condition	$PD$ (mV)	$I_{sc}$ ( $\mu A$ )	$R$ ( $K\Omega$ )
Before vanadate	$-38.3 \pm 6.2$	$-31.2 \pm 5.4$	$1.2 \pm 0.1$
After vanadate	$-8.2 \pm 2.3$	$-9.0 \pm 3.0$	$0.9 \pm 0.1$
$\Delta(\%)$	$-75.1 \pm 5.3$	$-72.2 \pm 4.9$	$-21.5 \pm 4.6$
$P(\Delta=0)$	$P < 0.001$	$P < 0.001$	$P < 0.001$

TABLE III

EFFECT OF VANADATE ON  $Cl^-$  FLUXES ACROSS OUABAIN-TREATED BLADDERS IN  $Na^+$  ( $Cl^- + HCO_3^-$ ) BATHING MEDIUM

Mean values  $\pm$  S.E. ( $n = 5$ ) of the measured  $Cl^-$  fluxes and calculated net fluxes and flux ratios of five pairs of mated hemi-bladders in Table II. Experimental conditions given in Table II. Statistical definitions given in Table I. The value of  $Cl^-$  flux before vanadate is the steady-state level during the two 30 min sampling periods immediately before addition of vanadate; the value of  $Cl^-$  flux after vanadate is that obtained during the two sampling periods 90–150 min after the addition of vanadate (Ref. 7). Net  $Cl^-$  fluxes and flux ratios were calculated from the individual flux differences and flux ratios of the five pairs of mated hemi-bladders. Flux values are for  $1.5 \text{ cm}^2$  of tissue area.

Condition	M-S $Cl^-$ flux ( $\mu A$ )	S-M $Cl^-$ flux ( $\mu A$ )	Net M-S $Cl^-$ flux ( $\mu A$ )	Flux ratio (M-S/S-M)
Before vanadate	$27.8 \pm 4.3$	$2.6 \pm 0.4$	$25.1 \pm 5.8$	$10.1 \pm 2.0$
After vanadate	$13.0 \pm 2.8$	$4.6 \pm 0.5$	$8.0 \pm 3.6$	$2.5 \pm 0.7$
$\Delta(\%)$	$-53.7 \pm 4.8$	$+78.2 \pm 15.9$	$-70.8 \pm 6.8$	$-73.8 \pm 5.0$
$P(\Delta=0)$	$P < 0.001$	$P < 0.001$	$P < 0.001$	$P < 0.001$

changes in  $Cl^-$  fluxes were checked in four pairs of mated hemi-bladders. While the M-to-S fluxes in the vanadate-treated hemi-bladders decreased by 49.6 and 59.7%, those of the untreated hemi-bladders decreased by 2.5 and 7.4%, respectively. The increases in the S-to-M fluxes of treated bladders (78.5 and 39.1%) were similar to those of untreated bladders (33.7 and 80.7%, respectively).

Since net  $Cl^-$  reabsorption (Table III) could account for 80% of  $I_{sc}$  (Table II), the effect of vanadate on  $HCO_3^-$  reabsorption, defined as  $I_{sc}$  minus the net  $Cl^-$  flux [12, 14, 15], could not be accurately determined. Its effect on  $HCO_3^-$  transport was measured in 11 ouabain-treated bladders bathed by  $HCO_3^-$ -rich,  $Cl^-$ -free ( $Cl^-$  replaced by  $SO_4^{2-}$ ) media in which  $I_{sc}$  approximates the net reabsorption of  $HCO_3^-$  [7, 14, 18]. In the presence of 1 mM vanadate,  $I_{sc}$  ( $16.2 \pm 2.2 \mu A$ ) declined  $76.1 \pm 16.3\%$  ( $n=5$ ) after 30 min and approached near-zero values at about 60 min; with 0.1 mM vanadate present,  $I_{sc}$  decreased  $37.8 \pm 9.2\%$  ( $n=6$ ) in 30 min and  $47.9 \pm 8.3\%$  ( $n=6$ )

in 60 min. The inhibition of the  $\text{HCO}_3^-$  moiety of  $I_{\text{sc}}$  predicts a potent inhibition of mucosal acidification [7, 19] by vanadate.

One additional effect of serosal vanadate on the electrical parameters of ouabain-treated bladders is noteworthy. In about 50% of the bladders studied, vanadate caused two damped oscillations in the electrical parameters superimposed on the more slowly declining base-line values of these parameters as shown in Fig. 1 (A); of the remaining bladders, 35% exhibited one fluctuation in  $PD$ ,  $I_{\text{sc}}$ , and  $R$ . Three or more fluctuations as shown in Fig. 1B were observed in three cases. These transients were observed under the following conditions: (i) 0.1 or 1 mM serosal vanadate; (ii) open-circuited or short-circuited bladders; (iii)  $\text{Cl}^-$ -rich or  $\text{Cl}^-$ -free media ( $\text{HCO}_3^-$  present); and (iv) in the presence of either 0 or 1 mM vanadate in the mucosal fluid.

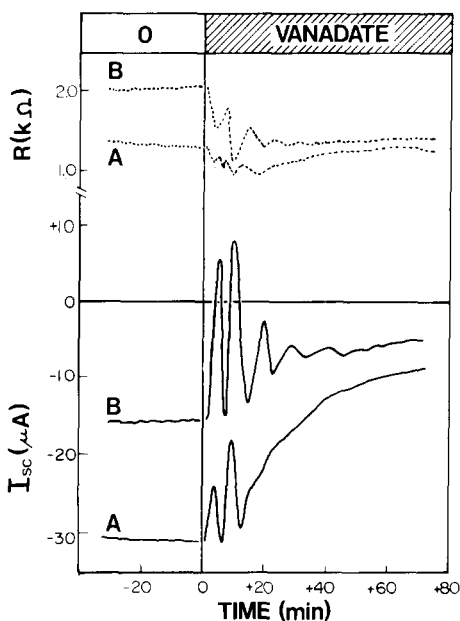


Fig. 1. Transient effect of vanadate on  $I_{\text{sc}}$  and  $R$  of ouabain-treated bladders in  $\text{Cl}^-$ -rich and  $\text{Cl}^-$ -free bathing media.  $R$  is shown in upper panel (dotted curves);  $I_{\text{sc}}$  is shown in lower panel (solid curves). Negative values of  $I_{\text{sc}}$  indicate that serosa is electronegative to mucosa. The time course of  $PD$  has been omitted for clarity but can be estimated from  $I_{\text{sc}} \times R$ . Data from two experiments (A and B) are shown. Experimental conditions: (A) 1 mM vanadate; bathing medium described in legend to Table II; (B) 0.1 mM vanadate; bathing medium as described in legend to Table II, except that  $\text{Cl}^-$  was replaced by  $\text{SO}_4^{2-}$  without changing the concentration of cations. Osmolality was readjusted with sucrose.

Whereas the rapid fall in  $PD$ ,  $I_{\text{sc}}$ , and  $R$  beginning 2 min after serosal vanadate addition may be caused, at least in part, by the onset of  $\text{H}_2\text{VO}_4^-$  diffusion across the basolateral membrane, the subsequent transients in the electrical parameters are not readily explained by such a process. Oscillations in ionic fluxes have been observed in mitochondria isolated from other tissues [20]. However, unlike the harmonic oscillations induced by various weak acidic anions in mitochondria, the oscillations induced by vanadate in the turtle bladder have a longer and progressively increasing period (from 4 to 12 min over a total time of about 50 min in Fig. 1). A tentative hypothe-

sis is that a vanadate-induced hyperpolarization of the basolateral membrane, in part, causes changes in the conductive flows of ions other than vanadate across this membrane.

It is concluded that vanadate inhibits  $\text{Na}^+$ ,  $\text{Cl}^-$ , and  $\text{HCO}_3^-$  reabsorption in the turtle bladder with a relative potency of  $\text{HCO}_3^- > \text{Cl}^- \sim \text{Na}^+$  transport. These data constitute the first demonstration of an inhibitory effect by vanadate on ouabain-insensitive epithelial anion transport. Earlier studies have shown that anion transport in the turtle bladder is not only ouabain-insensitive, but also  $\text{Na}^+$ -independent [9, 15]. Consequently, inhibition of electrolyte transport by vanadate is not restricted to a primary inhibition of cation transport, as has been frequently assumed. The potency of vanadate in turtle bladders is about 1/10 that of the disulfonic stilbenes in inhibiting  $\text{Cl}^-$  and  $\text{HCO}_3^-$  reabsorption [6, 7] and about 1/1000 that of ouabain in inhibiting  $\text{Na}^+$  transport [15].

To account for the observed vanadate-induced inhibition of anion and  $\text{Na}^+$  transport the following is proposed. (i) Vanadate interacts with the anion-selective pathways (or carriers) in the basolateral membrane [6–8] to retard anion transport. Several factors might be involved, e.g., electrostatic repulsion and steric hindrance due to the complex structural and binding properties of this transition metal anion [5, 10, 21], although vanadate actions at other cellular sites [22, 23], including mitochondria, cannot be excluded. (ii) Upon penetrating via the anion paths, vanadate inhibits  $\text{Na}^+$  transport by binding to the  $(\text{Na}^+ + \text{K}^+)\text{-ATPase}$  [15] on the cytoplasmic side of the membrane in a manner analogous to that described by Cantley et al. for red cells [5].

The concentration of vanadate used in this study was much higher than that used by Balfour et al. [1] *in vivo*. Nevertheless, the results are consistent with the notion that a vanadate-induced inhibition of active anion transport in the distal nephron segments [24] contributes to the potent diuretic action of this substance [1].

This study was supported by grants from the Ohio University Research Committee and the College of Osteopathic Medicine. Acknowledgement is due to Susan R. Ehrenspeck and James D. Cummings for their excellent technical assistance.

## References

- 1 Balfour, W.E., Grantham, J.J. and Glynn, I.M. (1978) *Nature* 275, 768
- 2 Cantley, L.C., Josephson, L., Warner, R., Yanagisawa, M., Lechene, C. and Guidotti, G. (1977) *J. Biol. Chem.* 252, 7421–7423
- 3 Grantham, J.J. and Glynn, I.M. (1979) *Am. J. Physiol.* 236, F530–F535
- 4 De Sousa, R.C. and Grosso, A. (1979) *Nature* 279, 803–804
- 5 Cantley, L.C., Resh, M.E. and Guidotti, G. (1978) *Nature* 272, 553–554
- 6 Ehrenspeck, G. and Brodsky, W.A. (1975) *Biochim. Biophys. Acta* 419, 555–558
- 7 Brodsky, W.A., Durham, J. and Ehrenspeck, G. (1979) *J. Physiol.* 287, 559–573
- 8 Brodsky, W.A., Cabantchik, Z.I., Davidson, N., Ehrenspeck, G., Kinne-Saffran, E.M. and Kinne, R. (1979) *Biochim. Biophys. Acta* 556, 490–508
- 9 Gonzalez, C.F., Shamoo, Y.F. and Brodsky, W.A. (1967) *Am. J. Physiol.* 212, 641–650
- 10 Kepert, D.L. (1973) in *Comprehensive Inorganic Chemistry* (Bailar, J.C., Jr., Emeleus, H.J., Nyholm, R. and Trotman-Dickenson, A.F., eds.), Vol. 4, pp. 607–672, Pergamon Press, Oxford
- 11 Gonzalez, C.F., Shamoo, Y.E. and Brodsky, W.A. (1969) *Biochim. Biophys. Acta* 193, 403–418
- 12 Gonzalez, C.F., Shamoo, Y.E., Wyssbrod, H.R., Solinger, R.D. and Brodsky, W.A. (1967) *Am. J. Physiol.* 213, 333–340

- 13 Steinmetz, P.R., Omachi, R.S. and Frazier, H.S. (1967) *J. Clin. Invest.* 46, 1541—1548
- 14 Gonzalez, C.F. (1969) *Biochim. Biophys. Acta* 193, 146—158
- 15 Solinger, R.S., Gonzalez, C.F., Shamoo, Y.E., Wyssbrod, H.R. and Brodsky, W.A. (1968) *Am. J. Physiol.* 215, 249—260
- 16 Schwartz, J.H. (1976) *Am. J. Physiol.* 231, 565—572
- 17 Leslie, B.R., Schwartz, J.H. and Steinmetz, P.R. (1973) *J. Physiol.* 225, 610—617
- 18 Gonzalez, C.F. and Schilb, T.P. (1969) *Biochim. Biophys. Acta* 193, 419—429
- 19 Brodsky, W.A. and Schilb, T.P. (1974) in *Current Topics in Membranes and Transport* (Bronner, F. and Kleinzeller, A., eds.), Vol. 5, pp. 161—224, Academic Press, New York
- 20 Gooch, V.D. and Packer, L. (1974) *Biochim. Biophys. Acta* 346, 245—260
- 21 Van Etten, R.L., Waymack, P.O. and Rehkop, D.M. (1974) *J. Am. Chem. Soc.* 96, 6782—6785
- 22 O'Neal, S.G., Rhoads, D.B. and Racker, E. (1979) *Biochem. Biophys. Res. Commun.* 89, 845—849
- 23 Simons, T.J.B. (1979) *Nature* 281, 337—338
- 24 Burg, M.B. (1976) *Kidney Int.* 9, 189—197